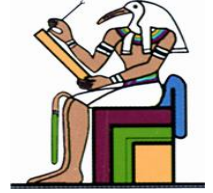




كلية الطب البيطري  
جامعة القاهرة

## وحدة ضمان الجودة



جامعة القاهرة

### Curriculum Vitae

#### **personal Information**

<b>Name</b>	<b>Manal Ahmed Farid Mohammed</b>	<b>optional photo</b>
<b>Title</b>	Department of Zoonoses, Faculty of Veterinary Medicine, Faculty of Veterinary Medicine, Cairo University.	
<b>Date of birth</b>		
<b>Place of birth</b>	Cairo Governorate, Egypt.	
<b>Citizenship</b>	Egyptian	

#### **Contact Information**

<b>Home phone</b>	
<b>Work phone</b>	35720399
<b>Mobile phone</b>	01023551081
<b>E-mail (s)</b>	<a href="mailto:manal.farid@staff.cu.edu.eg">manal.farid@staff.cu.edu.eg</a>
<b>Web site (s)</b>	-----
<b>Current Address</b>	

#### **Educational Qualifications**

<b>Ph.D.</b> Faculty of Vet. Med. Cairo University , 2011
<b>M.V.Sc.</b> Faculty of Vet. Med. Cairo University, 2006.
<b>B.V.Sc.</b> Faculty of Vet. Med. Cairo University May, 2002.

#### **Academic Positions**

- Lecturer of Vet. zoonoses, Faculty of Vet. Med. Cairo University 2012.

**University Lecturer:** (Spring Term, 2012)

School of Veterinary Medicine, University of Liverpool, UK



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-Teaching veterinary epidemiology for 2<sup>nd</sup> year undergraduate veterinary students in order to clarify the importance of epidemiology and statistics, both for veterinary science and human medical science and applying an epidemiological approach and basic epidemiological and statistical methods in appropriate situations.

- Assistant Lecturer of Zoonoses, Faculty of Vet. Med. Cairo University 2006.

- Demonstrator of Zoonoses, Faculty of Vet. Med. Cairo University 2002.

### **Thesis Title**

**Ph.D.** Studies on zoonotic Japanese encephalitis virus Muar strain

**M.V.Sc.** Possible Role of Laboratory Animals in Transmitting Some Occupational zoonoses

### **Areas of experience**

Molecular biotechnology; RNA extraction, RT PCR, PCR, primers design (manually and by using software), gene cloning and sequencing.

-Tissue culture; (VERO cells, mosquito cells, mouse astrocytes and human microglia)

-Virology; CL2 and CL3 skills: virus culture and harvesting, plaque assays and plaque reduction neutralization tests;

evaluating the efficacy of new anti-viral therapy such as Intra venous immunoglobulins.

-Bioinformatics; sequence alignment using (BLAST, Vector NTI suite software package, clustalX and ClustalW), prediction of the RNA secondary structure and determination of the three dimensional structure using PYMOL.

-Phylogenetic analyses using MEGA4, PAUP\* and GARLI.

-Evolutionary analyses using BEAST and analyses of selection pressure using DataMonkey software.

### **Projects**

### **Awards**

### **Professional Qualifications**

### **Training courses**



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**MRCVS examination preparation course**, Easter Bush Veterinary Centre, Roslin, University of Edinburgh, UK;

(6 – 8 April 2011) where practical training on horses, farm animals and small animals was performed.

**-Workshop on Molecular Evolution**, Czech Republic, Europe (10 – 30 January 2010); organized by experts in Molecular evolution from USA, UK, Canada and Australia. I gained intensive training on molecular evolution software and I got the chance to meet with most of phylogenetic and evolutionary software developers for (PAUP, BEAST, ...). Furthermore, I gained very good knowledge in transcriptome analysis using RNAseq to detect mutation events including: novel splicing events, gene fusions, SNPs, or other specific coding mutations within the genome.

**-Laboratory animals training course**, Modules 1, 2, 3 and 4, University of Liverpool, UK (December 2009).

**-Postgraduate courses**, University of Liverpool, Graduate School\_one day event workshops;

Scientific Writing, Creativity in Research, Writing Targeted Grant Proposals, Communicating with confidence, Viva Survivor, Project Management in the Real World,

### Computer Skills

Obtained ICDL (International Computer Driving Licence) in 2007.

Excellent in MS office packages with basic skills in Unix/Linux.

Familiar with several phylogenetic and evolutionary analyses software.

### Language Skills

Native Arabic and fluent English (written and spoken) and little German.

- I passed the International English Language Testing System (IELTS) in October 2010 with score 7 from University of Liverpool, UK.

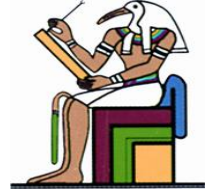
- I passed the Test of English as a Foreign Language (TOEFL) in 2007 with score 550 from AMIDEAST, Cairo, Egypt.

- I worked as Interpreter (Part Time) since October 2008 at Language Empire, North West, UK; facilitating effective Communication from English to Arabic and vice versa between clients in different areas, such as medical appointments, social services appointments and community events.



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Associate Fellow of the Higher Education Academy, UK since May 2012.

-Associate Member of Society for General Microbiology (SGM), UK since 2008.

-Member of veterinary surgeons syndicate in Egypt since 2002.

-I was accepted by Royal College Veterinary Surgeon (RCVS) as an overseas veterinary surgeon and I am allowed to see veterinary practice in UK. Also, I sat the RCVS Statutory Membership Examinations in May 2011 and I passed three subjects out of total four.

### **Other activities**

**Certified Trainer:** (September 2012 - present)

Faculty and leadership development center (FLDC).

I am responsible for teaching "Scientific Research Ethics" course.

**Reviewer:** (September 2012- Present)

Journal of Advanced Research (Elsevier).

**Senior Research Technician:** (April 2012 – September 2012)

University of Southampton , Faculty of Medicine, Infectious Disease Epidemiology Research Group.

-Processing clinical samples and microbial DNA for whole genome sequencing and microbial community analysis on **next generation DNA sequencing systems** and associated equipment.

-Ordering and maintaining laboratory supplies, reagents and equipment.

-liaising with suppliers to obtain quotations, purchase contracts, discounts and to arrange maintenance contracts.

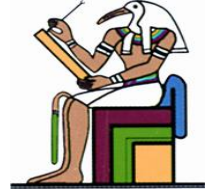
-Overseeing the efficient running of CL1 and CL2 culture facilities inclusive of ensuring that all equipment (primarily class I and II microbiological safety cabinets, and incubators) are maintained in a clean and sterile condition and to ensure regular supply of consumables.

-Ensuring all laboratory equipment is maintained in accordance with technical and health and safety procedures, diagnosing faults and liaising with suppliers to arrange replacements or on-site/off-site repairs. This will ensure minimal



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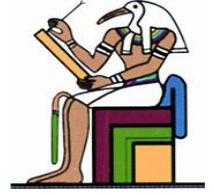
disruption to research work.

- Overseeing upkeep of carbon dioxide supplies and to ensure constant supply of carbon dioxide to culture incubators.
- Supporting and train staff in the use of, and to maintain, laboratory equipment e.g. DNA sequencing systems, PCR instrumentation, gel imagers, centrifuges.
- Creating and maintaining accurate logs of equipment usage for the purpose of aiding maintenance and where necessary for management of cost recovery from users.
- Helping manage any refurbishment process related to laboratories and offices occupied by staff and infrastructure.
- Monitoring the usage of facilities to ensure that appropriate codes of practice are followed, in order that a clean working environment is maintained.
- Assisting with all aspects of health and safety in laboratories in which staff carry out research activities, and actively implement appropriate changes to bench practices or local administration procedure as appropriate.
- Reporting accidents and health and safety breaches to the Divisional Health and Safety Officer.
- Assisting group members with the preparation of risk assessments and standard operating procedures for all experimental procedures, and to maintain records, to ensure compliance with relevant health and safety legislation.
- Assisting with health and safety inspections, to prepare reports of these, and follow up any actions required.
- Assisting with the induction of new staff/students, and to advise all group members on appropriate health and safety matters.



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### List of publications

**Mohammed, M. A.**, S. E. Galbraith, Alan D. Radford, Winifred Dove, Tomohiko Takasaki, Ichiro Kurane,

Tom Solomon (2011). "Molecular phylogenetic and evolutionary analyses of Muar strain of Japanese encephalitis virus reveal it is the missing fifth genotype." *Infection, Genetics and Evolution* 11 (5), 855-862.

**Mohammed, M. A.**, S. E. Galbraith, Alan D. Radford, Tom Solomon. (29<sup>th</sup> June 2010) The complete nucleotide and derived amino acid sequence of Muar strain of Japanese encephalitis virus was submitted into Genbank (accession no. HM596272.1).

**Mohammed, M. A.**, Siritorn Butrapet, Sareen E. Galbraith, Ichiro Kurane, Alan Radford & Tom Solomon. (27<sup>th</sup> April 2010) "Molecular Evolution of Japanese Encephalitis Virus", University of Liverpool, School of Infection and Host Defence Away Day, Ness Botanic Gardens Neston, Wirral. (Poster). This poster was ranked as the second best poster.

**Mohammed, M. A.**, Siritorn Butrapet, Sareen E. Galbraith, Ichiro Kurane, Alan Radford & Tom Solomon. (31<sup>st</sup> March 2010). "Molecular phylogenetic and evolutionary analyses of Muar strain of Japanese encephalitis virus: the missing 5th genotype". Society of General Microbiology (SGM) Spring 2010 Meeting, Edinburgh International Conference Centre. (Talk)

**Mohammed, M. A.**, Siritorn Butrapet, Tomohiko Takasaki, Ichiro Kurane, Sareen E. Galbraith & Tom Solomon. (1<sup>st</sup> April 2009) "Phylogenetic analysis of Muar strain of Japanese encephalitis virus reveals a possible 5th Genotype". Society of General Microbiology (SGM) Spring 2009 Meeting, Harrogate International Centre. (Talk)

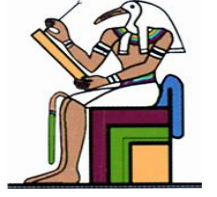
### Publications statistic

Journal's Publication		Conference's Publication		Authors			Total 5
Local	International	Local	International	Single	Shared		
					Internal	External --	
-	5		3				



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